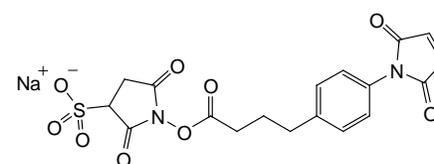
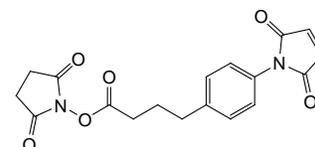


SMPB and Sulfo-SMPB

22416 22317

1762.2

Number	Description
22416	<p>SMPB (succinimidyl 4-[<i>p</i>-maleimidophenyl]butyrate), 50mg</p> <p>Molecular Weight: 356.33</p> <p>Spacer Arm Length: 11.6Å</p> <p>Storage: Upon receipt store desiccated at 4°C. Product is shipped at ambient temperature.</p>
22317	<p>Sulfo-SMPB (sulfosuccinimidyl 4-[<i>p</i>-maleimidophenyl]butyrate), 50mg</p> <p>Molecular Weight: 458.38</p> <p>Spacer Arm Length: 11.6Å</p> <p>Storage: Upon receipt store desiccated at -20°C. Product is shipped at ambient temperature.</p>



Introduction

SMPB and its water-soluble analog Sulfo-SMPB are heterobifunctional crosslinkers that contain *N*-hydroxysuccinimide (NHS) ester and maleimide groups that allow covalent conjugation of amine- and sulfhydryl-containing molecules. NHS esters react with primary amines at pH 7-9 to form amide bonds, while maleimides react with sulfhydryl groups at pH 6.5-7.5 to form stable thioether bonds. In aqueous solutions, hydrolytic degradation of the NHS ester is a competing reaction whose rate increases with pH. The maleimide group is more stable than the NHS-ester group but will slowly hydrolyze and also lose its reaction specificity for sulfhydryls at pH values > 7.5. For these reasons, conjugation experiments involving these crosslinkers are usually performed at pH 7.2-7.5, with the NHS-ester (amine-targeted) reaction being accomplished before or simultaneous with the maleimide (sulfhydryl-targeted) reaction.

SMPB and Sulfo-SMPB are often used to prepare antibody-enzyme and hapten-carrier protein conjugates in a two-step reaction scheme. First, the amine-containing protein is reacted with a several-fold molar excess of the crosslinker, followed by removal of excess (nonreacted) reagent by desalting or dialysis; finally, the sulfhydryl-containing molecule is added to react with the maleimide groups already attached to the first protein.

Sulfo-SMPB is soluble in water and many other aqueous buffers to approximately 10 mM, although solubility decreases with increasing salt concentration. SMPB is not directly water-soluble and must be dissolved first in an organic solvent such as dimethylsulfoxide (DMSO) or dimethylformamide (DMF); subsequent dilution into aqueous reaction buffer is generally possible, and most protein reactants will remain soluble if the final concentration of organic solvent is less than 10%.

Important Product Information

- SMPB and Sulfo-SMPB crosslinkers are moisture-sensitive. Store vial of reagent in desiccant at the specified temperature. Equilibrate vial to room temperature before opening to avoid moisture condensation inside the container. Dissolve needed amount of reagent and use it immediately before hydrolysis occurs. Discard any unused reconstituted reagent. Do not attempt to make and store stock solutions.
- Avoid buffers containing primary amines (e.g., Tris or glycine) and sulfhydryls during conjugation because they will compete with the intended reaction. If necessary, dialyze or desalt samples into an appropriate buffer such as phosphate buffered saline (PBS).

- Molecules to be reacted with the maleimide moiety must have free (reduced) sulfhydryls. Reduce peptide disulfide bonds with Immobilized TCEP Disulfide Reducing Gel (Product No. 77712). Reduce disulfide bonds in high molecular weight proteins using 5mM TCEP (1:100 dilution of Bond-Breaker™ TCEP Solution, Product No. 77720) for 30 minutes at room temperature, followed by two passes through an appropriate desalting column (e.g., Zeba™ Desalt Spin Columns). Be aware that proteins (e.g., antibodies) may be inactivated by complete reduction of disulfide bonds they contain. Selective reduction of hinge-region disulfide bonds in IgG may be accomplished with 2-Mercaptoethylamine•HCl (2-MEA, Product No. 20408). Sulfhydryls may be added to molecules using *N*-succinimidyl *S*-acetylthioacetate (SATA, Product No. 26102) or 2-iminothiolane•HCl (Traut's Reagent, Product No. 26101), which modify primary amines.

Procedure for Two-step Protein Crosslinking

Generally, a 10- to 50-fold molar excess of crosslinker over the amount of amine-containing protein results in sufficient maleimide activation to enable several sulfhydryl-containing proteins to be conjugated to each amine-containing protein. More dilute protein solutions require greater fold molar excess of reagent to achieve the same level of activation. Empirical testing is necessary to determine activation levels and final conjugation ratios that are optimal for the intended application.

A. Material Preparation

- Conjugation Buffer: Phosphate buffered saline (PBS, pH 7.2; e.g., Product No. 28372) or other amine- and sulfhydryl-free buffer at pH 6.5-7.5 (see Important Product Information) – adding EDTA to 1-5mM helps to chelate divalent metals, thereby preventing disulfide formation in the sulfhydryl-containing protein
- Desalting column to separate modified protein from excess crosslinker and reaction byproducts (e.g., Zeba™ Desalt Spin Columns)
- Amine-containing protein (Protein-NH₂) and sulfhydryl-containing protein (Protein-SH) to be conjugated

B. Protocol

Note: For best results, ensure that Protein-SH is prepared (see Important Product Information) and ready to combine with Protein-NH₂ in step 5.

1. Dissolve Protein-NH₂ in Conjugation Buffer at 0.1 mM (e.g., 5mg in 1mL for a 50kDa protein).
2. Add crosslinker to dissolved Protein-NH₂ at 1mM final (= 10-fold molar excess):
 - For Sulfo-SMPB, dissolve 4.58mg Sulfo-SMPB in 1mL of water or diluted (low-salt) Conjugation Buffer (makes 10mM temporary stock) and immediately add 100µL of this solution to 1mL of Protein-NH₂ solution.
 - For SMPB, dissolve 3.55mg SMPB in 1mL DMSO (makes 10mM) and then add 100µL of this solution to 1mL of Protein-NH₂ solution.
3. Incubate reaction mixture for 30 minutes at room temperature or 2 hours at 4°C.
4. Remove excess crosslinker using a desalting column equilibrated with Conjugation Buffer.

Note: Follow the desalting column product instructions to determine which fractions contain Protein-NH₂. Alternatively, locate the protein by measuring for fractions having peak absorbance at 280nm; however, be aware that the NHS-ester leaving group also absorbs strongly at 280nm.

5. Combine and mix Protein-SH and desalted Protein-NH₂ in a molar ratio corresponding to that desired for the final conjugate and consistent with the relative number of sulfhydryl and activated amines that exist on the two proteins.
6. Incubate the reaction mixture at room temperature for 30 minutes or 2 hours at 4°C.

Note: Generally, there is no harm in allowing the reaction to proceed for several hours or overnight, although usually the reaction will be complete in the specified time. To stop the conjugation reaction before completion, add buffer containing reduced cysteine at a concentration several times greater than the sulfhydryls of Protein-SH.

Note: Conjugation efficiency may be estimated by electrophoresis separation and subsequent protein staining.

Related Thermo Scientific Products

Table 1. Noncleavable NHS/Maleimide crosslinkers.

Crosslinker	Name	Spacer Arm Length (Å)	Spacer Arm Composition (between ester and maleimide)	Product No. (NHS)	Product No. (Sulfo-NHS)
	AMAS	4.4	Alkane	22295	NA
	BMPS	5.9	Alkane	22298	NA
	GMBS	7.3	Alkane	22309	22324
	MBS	7.3	Aromatic	22311	22312
	SMCC	8.3	Cyclohexane	22360	22322
	EMCS	9.4	Alkane	22308	22307
	SMPB	11.6	Alkane/Aromatic	22416	22317
	SMPH	14.2	Alkane/Amide	22363	NA
	LC-SMCC	16.2	Alkane/Amide/Cyclohexane	22362	NA
	KMUS	16.3	Alkane	NA	21111

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