pUC19 DNA/MspI (HpaII) Marker, 23

Catalog Number SM0221, SM0222

Pub. No. MAN0012992 **Rev.** C.00

WARNING! Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves. Safety Data Sheets (SDSs) are available from **thermofisher.com/support.**

Contents and storage

Cat. No.	Contents	Amount	Storage
SM0221	pUC19 DNA/MspI (HpaII) Marker, 23	50 µg (for 100 applications), 0.5 µg/µL	-25 °C to -15 °C
	6X DNA Loading Dye	1 mL	
SM0222	pUC19 DNA/MspI (HpaII) Marker, 23	250 (5 x 50) µg (for 500 applications), 0.5 µg/µL	
	6X DNA Loading Dye	2 x 1 mL	

Description

pUC19 DNA was completely digested with MspI, purified and dissolved in a storage buffer.
The DNA Marker contains the following 13 discrete fragments (in base pairs): 501, 489, 404, 331, 242, 190, 147, 111, 110, 67, 34, 34, 26.

Storage Buffer

10 mM Tris-HCl (pH 7.6), 1 mM EDTA.

6X DNA Loading Dye

10 mM Tris-HCl (pH 7.6), 0.03 % bromophenol blue, 0.03 % xylene cyanol FF, 60 % glycerol and 60 mM EDTA.

Protocol for Loading

Loading mixture for the 5 mm gel lane*:

Components	Gels	
	Agarose	**Polyacrylamide**
DNA ladder (0.5-1 µg)	1-2 µL	1-2 µL
6X DNA Loading Dye	1 µL	0.5 µL
Deionized water	4-3 µL	1.5-0.5 µL
	6 µL	3 µL

Step 1: Mix gently
Step 2: Load on the gel

*For gels with other lane widths, the components of the mixture should be scaled either up or down. Use 0.2-0.4 µL (0.1-0.2 µg) of DNA ladder per 1 mm of lane.

Recommendations

- Do not heat before loading
- Dilute your DNA sample with the 6X DNA Loading Dye (#R0611, supplied with the ladder): mix 1 volume of the dye solution with 5 volumes of the DNA sample;
- For DNA band visualization with SYBR™ Green and other intercalating dyes, do not add the dyes into the sample, use gel staining after electrophoresis or include dyes into agarose gel to avoid aberrant DNA migration.
- **Important note:** For DNA bands visualization with GelRed™ use gel staining after electrophoresis to avoid aberrant DNA migration.

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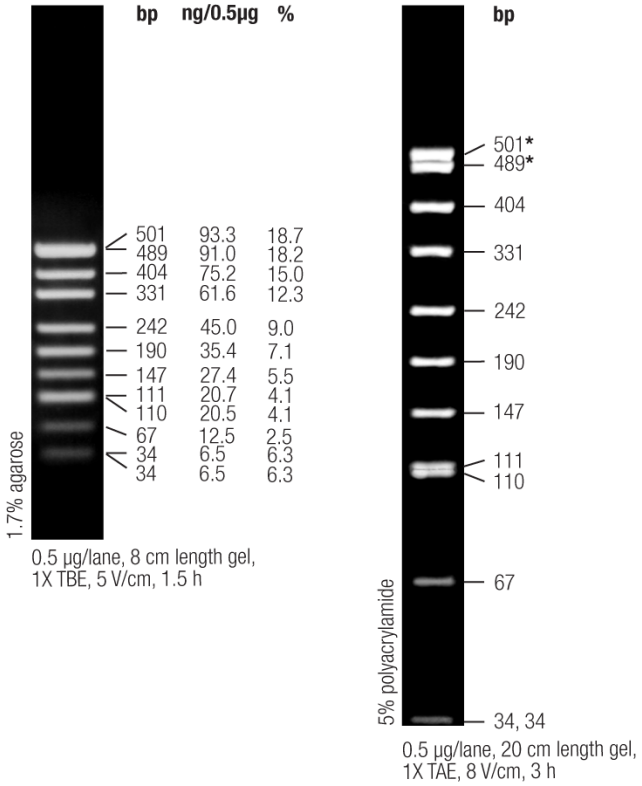

*The 501 and 489 bp bands migrate anomalously (1, 2, 3).
26 bp fragment is not visible and comprises 1.0 %

References

1. Stellwagen, N.C., Anomalous electrophoresis of deoxyribonucleic acid restriction fragments on polyacrylamide gels, Biochemistry, 22, 6186-6193, 1983.
2. Lane, D., et al., Use of gel ratardation to analyze protein – nucleic acid interactions, Microbiological Reviews, 56, 509-528, 1992.
3. Stellwagen, N.C., Conformational isomers of curved DNA molecules can be observed by polyacrylamide gel electrophoresis, Electrophoresis, 21, 2327-2334, 2000.

Limited product warranty

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