PureLink[®] HiPure Expi Megaprep Kit



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	Package	Catalog Numbers	Amount:
	Contents	K210008XP	4 preps
	Storage Conditions	 Store all components at room temperature. 	
	Required Materials	 Vacuum source equipped with regulator (capable of-600 to-800 mbar) Appropriately sized tubes and bottles 1000-mL Stericup® Receiver flask 250-mL Stericup® Receiver flask Centrifuge and rotor capable of >12,000 x g at 4°C 	
	Timing	Bacterial culture: overnight Purification: 90 minutes	
Å	Selection Guide	Go online to view related products: PureLink® Nucleic Acid Purification Kits Expi293™ Expression System	
	Product Description	 large quantities of trans exchange resin. The kit without centrifugation i The PureLink® HiPure E ultrapure plasmid DNA bacterial culture. High Yield–Isolate over purification using 1 L of 	Expi Megaprep Kit typically isolates 4 mg of high quality, with inherently low endotoxin levels from 500 mL of 5 mg of high quality plasmid DNA from a single Exacterial culture volume. levels $(0.1-1.0 \text{ EU}/\mu g)$, and A260/280 >1.8, making it ideal
	Important Guidelines	 label). Indicate that RNa If precipitate is observed bath until the solution of Grow transformed <i>E. co</i> or 2.5 L (low copy numl Do not over-dry DNA. I 	suspension Buffer (R3) and mix well (see instructions on ase A has been added on the bottle label. Store at 4°C. d in the Lysis Buffer (L7), warm the buffer in a 37°C water lears. Swirl contents gently to resuspend. <i>li</i> in LB medium. Use 500 mL (high copy number plasmid) per plasmid) of an overnight culture. If the DNA pellet is difficult to resuspend, allow the pellet for a longer period of time.
	Online Resources	Visit our product page for information and protocols visit www.lifetechnologies	. For support,

For Research Use Only. Not for use in diagnostic procedures.

Megaprep Plasmid Isolation Protocol

Steps		Procedure Details	
1	Harvest	1. Sediment cells by centrifugation at 4,000 × g for 15 min at 4°C. Discard all medium.	
2	Resuspend	2. Add 50 mL Resuspension Buffer (R3) with RNase A to the cell pellet and resuspend the pellet until it is homogeneous.	
3	Lyse	 Add 50 mL Lysis Buffer (L7). Mix gently by inverting the capped tube until the mixture is homogeneous. Do not vortex. Incubate at room temperature for 5 minutes. 	
4	Precipitate	4. Add 50 mL Precipitation Buffer (N3). Mix immediately by inverting the tube until the mixture is homogeneous. Do not vortex.	
5	Clarify	5. Pour the lysate into a lysate filtration cartridge attached to a receiver flask . Incubate for 2 minutes. Connect a vacuum source and filter the lysate.	
6	Wash	 Add 50 mL Wash Buffer (W8) to the filtration cartridge and gently stir precipitate with a spatula. Apply vacuum. The clarified lysate contains the plasmid DNA. 	
7	Equilibrate	 Add 100 mL Equilibration Buffer (EQ1) to a DNA-binding cartridge attached to a receiver flask. Connect a vacuum source and drain the cartridge. 	
8	Bind	 Load the clarified lysate (from step 6) onto the DNA-binding cartridge. Apply vacuum and drain solution. 	
9	Wash	 Add 175 mL Wash Buffer (W8) and apply vacuum. Repeat wash step. Attach DNA-binding cartridge to a new receiver flask. 	
10	Elute	 Add 50 mL Elution Buffer (E4) to the DNA-binding cartridge. Apply soft vacuum (-100 to -200 mbar) and draw 10-20 mL of solution. Stop the vacuum and incubate for 1 minute. Apply vacuum to all the liquid has passed from the cartridge. 	
11	Precipitate and Wash	11. Add 0.7 volume of isopropanol to the eluate. Mix well. Centrifuge at >12,000 × g for 30 minutes at 4°C. Remove and discard the supernatant. Wash the DNA pellet in 20 mL 70% ethanol. Centrifuge at >12,000 × g for 10 minutes at 4°C. Remove the supernatant.	
12	Resuspend	12. Air-dry the pellet for 10 minutes, then resuspend the purified plasmid DNA in TE Buffer (TE). Store plasmid DNA at -20°C.	

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